



First records of two genera and thirteen species of Tabanidae (Diptera) from Honduras

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Academic editor: T. Dikow | Received 25 October 2021 | Accepted 3 January 2022 | Published 26 January 2022

http://zoobank.org/3323F2E7-F3A8-49D1-9F46-21DABC882AF2

Citation: Veroy K, Orozco J, Henriques AL (2022) First records of two genera and thirteen species of Tabanidae (Diptera) from Honduras. ZooKeys 1084: 27–42. https://doi.org/10.3897/zookeys.1084.77038

Abstract

This works presents information on the diversity of the Tabanidae of Honduras as a product of the examination of 386 specimens and a literature review. Thirteen species and two genera (*Bolbodimyia* and *Dasychela*) are recorded from the country for the first time. Eighty-five species distributed in 22 genera, five tribes, and three subfamilies are now known from Honduras. A key to the subfamilies, tribes, and genera of the known Honduran species is also included. All new records are mapped and illustrated to aid in the identification of the species.

Keywords

Central America, diversity, horse flies, tabanids, taxonomy

Introduction

Tabanidae is a family of Diptera that includes flies considered of medical and veterinary importance due to the blood sucking habits of the adults. Currently the group contains around 4,400 species worldwide (Pape et al. 2011). The Neotropical region has the highest diversity, with approximately 1,205 species and about 28% of the global fauna (Henriques et al. 2012), but many its areas continue to be unexplored.

The best known tabanid faunas in Central America are those of Costa Rica and Panama thanks in big part to the works of Fairchild (1961), Hogue and Fairchild (1974), Fairchild (1986), and Burger (2002). Currently, 146 species of tabanids are known from Costa Rica (Borkent et al. 2018) and 152 from Panama (Fairchild 1986). For Honduras, few works deal with the diversity of horseflies in the country, i.e., Bequaert (1925), Root (1925), and James (1950). Coscarón and Papavero (2009), in their catalog for the neotropics, listed 70 species of Tabanidae from Honduras. Henriques (2016) added two additional species, *Scione maculipennis* (Schiner) and *Philipotabanus ebrius* (Osten Sacken), for a total of 72 species.

Honduran species diversity is poorly known for many groups. Linares and Orozco (2017) estimated that at least half of the insects in the country are known unknowns, species already described that are not recorded. This poor understanding of the diversity makes conducting ecological and conservation studies very difficult in the country.

This work presents for the first time an overview of the tabanids of Honduras. By nature, this is vastly incomplete as there are many more habitats to sample and collections to revise. In comparison, Costa Rica with less than half the size of Honduras has more than twice the number of known species of tabanids. The aims of this article are: 1) to present the new findings regarding the species diversity in the country, 2) to integrate the records on the tabanid fauna of Honduras scattered in the literature, 3) to provide an updated list of the species, and 4) to create a key for the genera of tabanids known in the country.

Methods

Material of Tabanidae deposited at the Insect Collection at Zamorano University (EAPZ) (Zamorano, Honduras) was examined. Fieldwork was done using H-traps (Egri et al. 2013), light traps, and an aerial net in several locations in Honduras. Specimens were studied under a Leica EZ4 stereo microscope using the keys provided by Bequaert (1931), Philip (1954), Fairchild and Philip (1960), Fairchild (1976), Wilkerson (1979), Fairchild (1983, 1986), Fairchild and Wilkerson (1986), Coscarón and González (1991), Burger (1996), Henriques (2006), Krolow et al. (2007), Burger (2009), Krolow and Henriques (2010), Turcatel et al. (2010), Carmo and Henriques (2019), and Turcatel (2019).

Distributional records were obtained from label data and from the literature.

A species distribution map was made for the new records using SimpleMappr (https://www.simplemappr.net/) and Microsoft Power Point v. 2112.

Photographs were taken using a Canon 100 mm lens mounted on a Canon Rebel T5i attached to a macro rail. Composite images were obtained using PICOLAY v. 2020–02–06 (http://www.picolay.de). Individual images were organized in plates in GIMP v. 2.10.24 (http://www.gimp.org).

Results and discussion

Bolbodimyia philipi Stone, 1954

E.M. Bermúdez & Villalba, 1989

Catachlorops baliopterus Gorayeb, L. Bemúdez,

Eighteen genera and 47 species were found in the 386 specimens examined. Thirteen species and two genera are recorded for the first time (Fig. 1).

With these new records Honduras has now a diversity of 85 species of horseflies (Table 1). This represents an increase of 15.3% compared to the previously known taxa (72 species) but it's still a low number, and many more species are expected to be discovered in the future. Two additional species, *Tabanus femoralis* Kröber from Escuela Agricola Panamericana Zamorano, Francisco Morazan, and *Stypommisa lerida* (Fairchild) from 15 km west of La Ceiba, Atlántida, are recorded in GBIF (https://www.gbif.org/es/occurrence/3048772282 and https://www.gbif.org/es/occurrence/3385753663). Since this material was not examined, it is not included in the list, but the records are probably valid.

Table 1. Species of Tabanidae from Honduras. Distributions according to Coscarón and Papavero (2009), except were indicated.

Taxon	Distribution
CHRYSOPSINAE	
CHRYSOPSINI	
Chrysops soror Kröber, 1925	Guatemala, Belize, Honduras, Costa Rica, Panama, Colombia, Venezuela
Chrysops auroguttatus Kröber, 1930	Mexico to Colombia
Chrysops latifasciatus Bellardi, 1859	Mexico to Nicaragua
Chrysops melaenus Hine, 1925	Honduras (new record), Nicaragua, Costa Rica to Venezuela
Chrysops mexicanus Kröber, 1926	Mexico to Colombia
Chrysops pachycnemius Hine, 1905	Mexico to Honduras
Chrysops scalaratus Bellardi, 1859	Mexico to Panama
Chrysops variegatus (De Geer, 1776)	Mexico to Argentina
Chrysops willistoni Hine, 1925	Mexico to Honduras
Silvius melanopterus (Hine, 1905)	Mexico to Honduras
PANGONIINAE	
PANGONIINI	
Esenbeckia illota (Williston, 1901)	Mexico to Honduras
Esenbeckia mejiai Fairchild, 1942	Guatemala to Costa Rica
Esenbeckia prasiniventris (Kröber, 1929)	Guatemala to Ecuador and Trinidad, Brazil
Esenbeckia translucens (Macquart, 1846)	Mexico to Peru and Brazil
Esenbeckia wiedemanni (Bellardi, 1859)	Mexico, Honduras (new record)
SCIONINI	
Fidena flavipennis Kröber, 1931	Mexico to Venezuela
Fidena rhinophora (Bellardi, 1859)	Mexico to Venezuela and Peru
Scione aurulans (Wiedemann, 1830)	Mexico to Costa Rica
Scione maculipennis (Schiner, 1868)	Honduras, Costa Rica to Venezuela, Ecuador*
TABANINAE	
DIACHLORINI	
Bolbodimyia atrata (Hine, 1904)	USA, Mexico, Honduras (new record)
Bolbodimyia erythrocephala (Bigot, 1892)	Honduras (new record), Costa Rica, Panama, Ecuador
Bolbodimyia galindoi Fairchild, 1964	Honduras (new record), Costa Rica to Colombia

Guatemala, El Salvador, Honduras (new record), Costa Rica, Panama, Colombia

Mexico, Honduras, Costa Rica

Tabanus oculus Walker, 1848

Tabanus picturatus Kröber, 1931

Tabanus pruinosus Bigot, 1892

Tabanus polyphemus Fairchild, 1958

Tabanus pseudoculus Fairchild, 1942

Distribution Taxon Catachlorops fulmineus (Hine, 1920) Honduras to Panama, Colombia, Ecuador Catachlorops scurrus (Fairchild, 1958) Mexico to Panama Chlorotabanus inanis (Fabricius, 1787) Mexico to Peru and Brazil Mexico to Ecuador, Brazil, Trinidad Chlorotabanus mexicanus (Linnaeus, 1758) Dasychela badia (Kröber, 1931) Honduras (new record), Costa Rica, Panama Diachlorus ferrugatus (Fabricius, 1805) USA to Costa Rica, Bahamas Islands Dichelacera costaricana (Fairchild, 1941) Honduras, Costa Rica Dichelacera grandis Philip, 1943 Guatemala, Belize, Honduras Dichelacera marginata Macquart, 1847 Honduras (New record), Nicaragua to Brazil and Peru Dichelacera pulchroides Fairchild & Philip, 1960 Mexico, Honduras Dichelacera regina Fairchild, 1940 Honduras to Ecuador Dichelacera scapularis Macquart, 1847 Mexico to Panama Dichelacera submarginata Lutz, 1915 Honduras (new record), Costa Rica to Venezuela, Peru, Bolivia Lepiselaga crassipes (Fabricius, 1805) Mexico to Argentina Leucotabanus exaestuans (Linnaeus, 1758) Mexico to Bolivia, Argentina, and Trinidad Leucotabanus nigriventris Kröber, 1931 Mexico to Panama Phaeotabanus longiappendiculatus Mexico to Panama (Macquart, 1855) Philipotabanus ebrius (Osten Sacken, 1886) Honduras, Costa Rica, Panama* Philipotabanus elviae (Fairchild, 1943) Honduras (new record), Costa Rica, Panama Philipotabanus kompi (Fairchild, 1943) Belize, Honduras Philipotabanus magnificus (Kröber, 1934) Honduras to Venezuela and Ecuador Philipotabanus nigrinubilus (Fairchild, 1953) Honduras, Costa Rica, Panama, Colombia, Ecuador Philipotabanus plenus (Hine, 1907) Guatemala to Colombia Rhabdotylus venenatum (Osten Sacken, 1886) Guatemala to Ecuador Selasoma tibiale (Fabricius, 1805) Mexico to Argentina Stenotabanus fulvistriatus (Hine, 1912) Mexico to Panama Stenotabanus littoreus (Hine, 1907) Mexico to Panama Honduras, Costa Rica, Panama, Trinidad, Venezuela. Stenotabanus maculifrons (Hine, 1907) Stibasoma chionostigma (Osten Sacken, 1886) Mexico to Colombia Stibasoma flaviventris (Macquart, 1848) Mexico to Brazil Stibasoma panamense Curran, 1934 Honduras to Ecuador and Venezuela Stypommisa captiroptera (Kröber, 1930) Mexico to Guyana, Brazil, Paraguay Stypommisa changena Fairchild, 1986 Honduras (new record), Costa Rica, Panama Mexico, Guatemala, Honduras Stypommisa u-nigrum Philip, 1977 **TABANINI** Poeciloderas quadripunctatus (Fabricius, 1805) Mexico to Argentina Tabanus abattenuis Philip, 1969 Mexico, Guatemala, El Salvador, Honduras, Nicaragua Tabanus bigoti Bellardi, 1859 Mexico to Colombia and Venezuela Tabanus claripennis (Bigot, 1892) Honduras (new record), West Indies, Costa Rica to Paraguay, Brazil, Argentina, and Chile Tabanus colombensis Macquart, 1846 USA to Trinidad, Venezuela, Ecuador, Brazil Mexico to Venezuela, Hispaniola, Trinidad, Martinique Tabanus commixtus Walker, 1860 Tabanus defilippii Bellardi, 1859 Mexico to Panama Tabanus dorsifer Walker, 1860 USA, Mexico, Honduras Tabanus erebus Osten Sacken, 1886 Honduras, Nicaragua, Costa Rica, Panama Tabanus jilamensis Hine, 1925 Honduras Tabanus morbosus Stone, 1938 USA, Mexico to Panama Tabanus nebulosus De Geer, 1776 Belize, Honduras (New record), Costa Rica, Trinidad, Barbados to Brazil and Argentina Tabanus occidentalis Linnaeus, 1758 Mexico to Argentina, Trinidad

Mexico to Panama

Mexico, Belize, Honduras

Mexico to Colombia

USA to Panama

Guatemala to Colombia, Venezuela, Ecuador, and Trinidad

Taxon	Distribution
Tabanus pungens Wiedemann, 1828	USA, Neotropics (except West Indies and Chile), Trinidad
Tabanus quinquepunctatus Hine, 1925	Guatemala, Belize, Honduras, Costa Rica, Panama
Tabanus secundus Walker, 1848	Guatemala to Peru, Surinam, and Paraguay
Tabanus subruber Bellardi, 1859	Mexico, Guatemala, Honduras
Tabanus unipunctatus (Bigot, 1892)	Mexico to Colombia
Tabanus unistriatus Hine, 1906	Guatemala to Ecuador
Tabanus vittiger ssp. guatemalanus Hine, 1906	USA, Bahamas, West Indies, Mexico to Surinam, French Guiana, and Brazil
Tabanus xenorhynchus Fairchild, 1947	Guatemala to Panama
Tabanus yucatanus Townsend, 1897	Mexico, Guatemala, El Salvador, Honduras, Nicaragua

^{*} Distribution according to Henriques (2016).

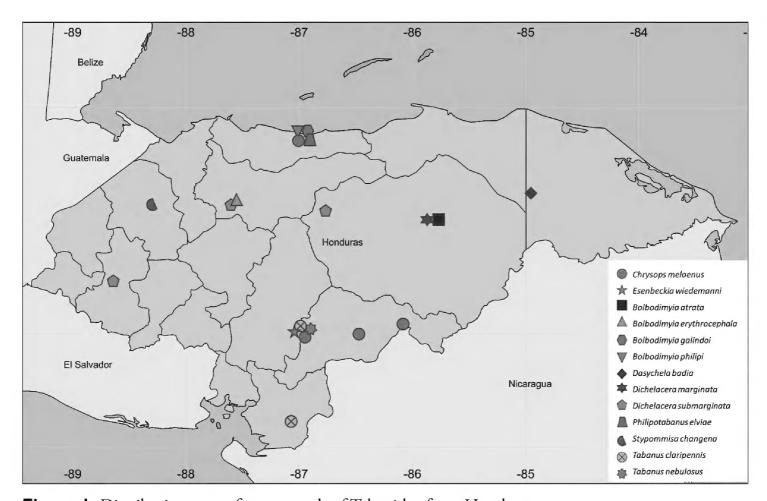


Figure 1. Distribution map of new records of Tabanidae from Honduras.

New Tabanidae from Honduras

CHRYSOPSINAE CHRYSOPSINI

Chrysops melaenus Hine, 1925

Figure 2A

Distribution. Previously known from Nicaragua to Venezuela (Coscarón and Papavero 2009).

Material examined. Honduras: 1♂, Atlántida, RVS Cuero y Salado, Salado Barra, 15°46′02″N, 86°59′51″W, 2 m, 25.i.2000, R. Cave, R. Cordero and J. Torres leg.; EAPZ22.445. 1♂, El Paraíso, 5.3 km N Cifuentes, 14°05′48″N, 86°06′57″W, 13.vi.1999, R. Cave and J. Torres leg.; EAPZ69.749. 1♀, El Paraíso, Danlí, Cerro

Apaguiz 14°00'27"N, 86°32'26"W, 20.ii.1988, R. Cordero leg.; EAPZ42.723. 1♀, Francisco Morazán, 32 km Tegucigalpa, El Zamorano, 14°01'N, 87°00'W, J. Cabezas leg.; EAPZ42.698.

PANGONIINAE PANGONIINI

Esenbeckia wiedemanni (Bellardi, 1859)

Figure 2B, C

Distribution. Previously known exclusively from Mexico (Coscarón and Papavero 2009). **Material examined.** Honduras: 1♂, 1♀, Francisco Morazán, Masicarán, Uyúca, 14°01′00″N, 87°05′00″W, 10–15.xi.2016, E. van den Berghe leg.; EAPZ42.764.

TABANINAE DIACHLORINI

Bolbodimyia atrata (Hine, 1904)

Figure 2D

Distribution. Previously known from U.S.A. and Mexico (Coscarón and Papavero 2009). **Material examined.** Honduras: 233, Olancho, El Murmullo, Sierra de Agalta, 15°01'00"N, 85°47'00"W, 28.vi.1997, R. Cave leg.; EAPZ69.815.

Bolbodimyia erythrocephala (Bigot, 1892)

Figure 2E

Distribution. Previously known from Costa Rica, Panama, Ecuador (Coscarón and Papavero 2009), and Colombia (Wolff and Miranda-Esquivel 2016).

Material examined. Honduras: 1♀, Yoro, Par. Nac. Pico Pijol, 15°13′00″N, 87°33′00″W, 22–23.vi.1998, R. Cave leg.; EAPZ42.652.

Bolbodimyia galindoi Fairchild, 1964

Figure 3A, B

Distribution. Previously known from Costa Rica to Colombia (Coscarón and Papavero 2009).

Material examined. Honduras: 1♂, 1♀, Atlántida, Par. Nac. Pico Bonito, Rio Zacate, 15°41'35"N, 86°55'58"W, 35 m, 5.iii.2000, R. Cave, R. Cordero and J. Torres leg.; EAPZ27.180.

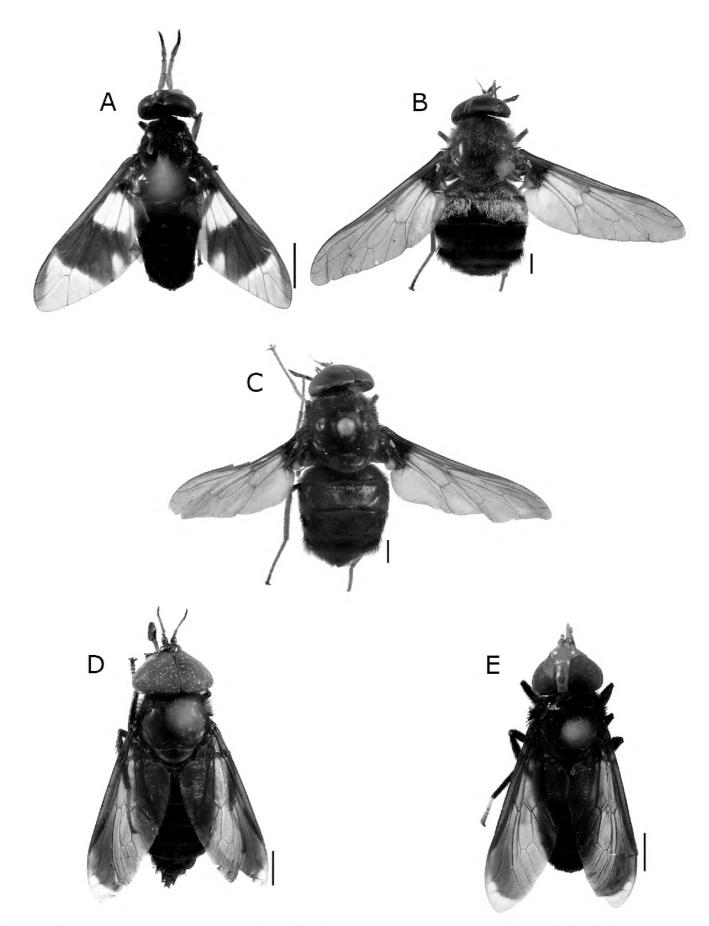


Figure 2. New records of Tabanidae from Honduras **A** *Chrysops melaenus* Hine (\updownarrow) **B, C** *Esenbeckia wiedemanni* (Bellardi) (\updownarrow , \eth) **D** *Bolbodimyia atrata* (Hine) (\eth) **E** *B. erythrocephala* (Bigot) (\updownarrow). Scale bars: 2 mm.

Bolbodimyia philipi Stone, 1954

Figure 3C

Distribution. Previously known from Guatemala, El Salvador, Costa Rica, Panama, and Colombia (Coscarón and Papavero 2009).

Material examined. Honduras: 1♂, Atlántida, Cuero y Salado, Salado Barra, 15°46′02″N, 86°59′51″W, 2 m, 25.i.2000, R. Cave, R. Cordero and J. Torres leg.; EAPZ22.452.

Dasychela badia (Kröber, 1931)

Figure 3D

Distribution. Previously known from Costa Rica and Panama (Coscarón and Papavero 2009).

Material examined. Honduras: 23♀♀, Gracias a Dios, Ciudad Blanca, 15°14′47″N, 84°58′2″W, 250 m, 15–26.ii.2017, E. van den Berghe leg., light trap; EAPZ43.577.

Dichelacera marginata Macquart, 1847

Figure 3E

Distribution. Previously known from Nicaragua to Brazil and Peru (Coscarón and Papavero 2009).

Material examined. Honduras: 1\(\text{?}\), Olancho, El Murmullo, Sierra de Agalta, 15°01'00"N, 85°47'00"W, 28.vi.1997, R. Cave leg.; EAPZ44.214.

Dichelacera submarginata Lutz, 1915

Figure 4A, B

Distribution. Previously known from Costa Rica to Venezuela, Peru, and Bolivia (Coscarón and Papavero 2009).

Material examined. Honduras: 1♀, Olancho, La Muralla, 15°04′56″N, 86°45′24″W, 26–30.iii.2013, O. Schlein leg.; EAPZ42.549. 1♂, Lempira, Par. Nac. Celaque, 14°28′46″N, 88°38′35″W, 1400 m, 27.iv.2018, E. van den Berghe leg.; EAPZ69.831. 1♂, Yoro, Par. Nac. Pico Pijol, Linda Vista, 15°10′35″N, 87°35′10″W, 1450 m, 21.iv.1999, R. Cave and J. Torres leg.; EAPZ42.829.

Philipotabanus elviae (Fairchild, 1943)

Figure 4C

Distribution. Previously known from Costa Rica and Panama (Coscarón and Papavero 2009).

Material examined. Honduras: $12 \circlearrowleft \circlearrowleft$, Atlántida, Par. Nac. Pico Bonito, Rio Zacate, $15^{\circ}41'35$ "N, $86^{\circ}55'58$ "W, 35 m, 5.v.2000, R. Cave leg.; EAPZ29.665.

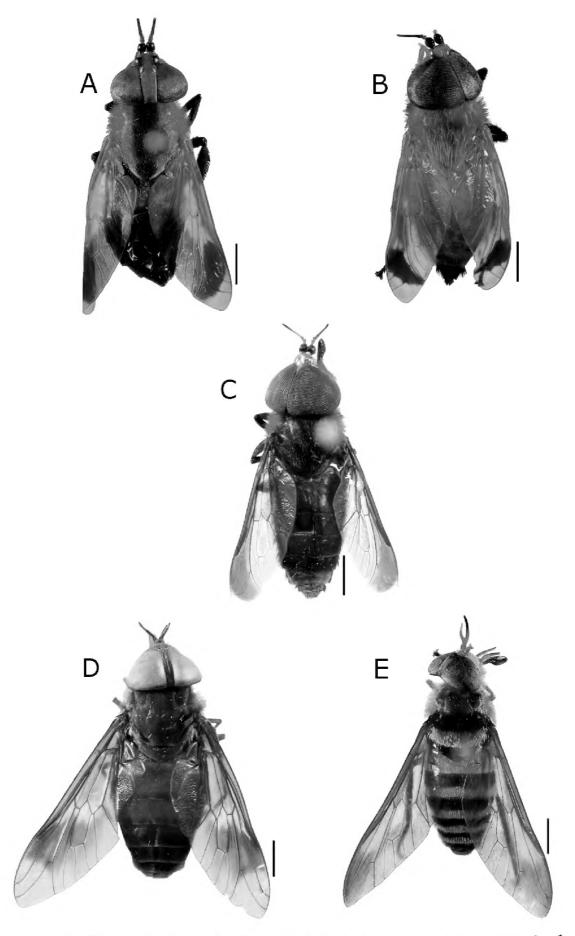


Figure 3. New records of Tabanidae from Honduras. **A, B** *Bolbodimyia galindoi* Fairchild $(\diamondsuit, \diamondsuit)$ **C** *B. philipi* Stone (\diamondsuit) **D** *Dasychela badia* (Kröber) (\diamondsuit) **E** *Dichelacera marginata* Macquart (\diamondsuit) . Scale bars: 2 mm.

Stypommisa changena Fairchild, 1986 Figure 4D

Distribution. Previously known from Costa Rica and Panama (Coscarón and Papavero 2009).

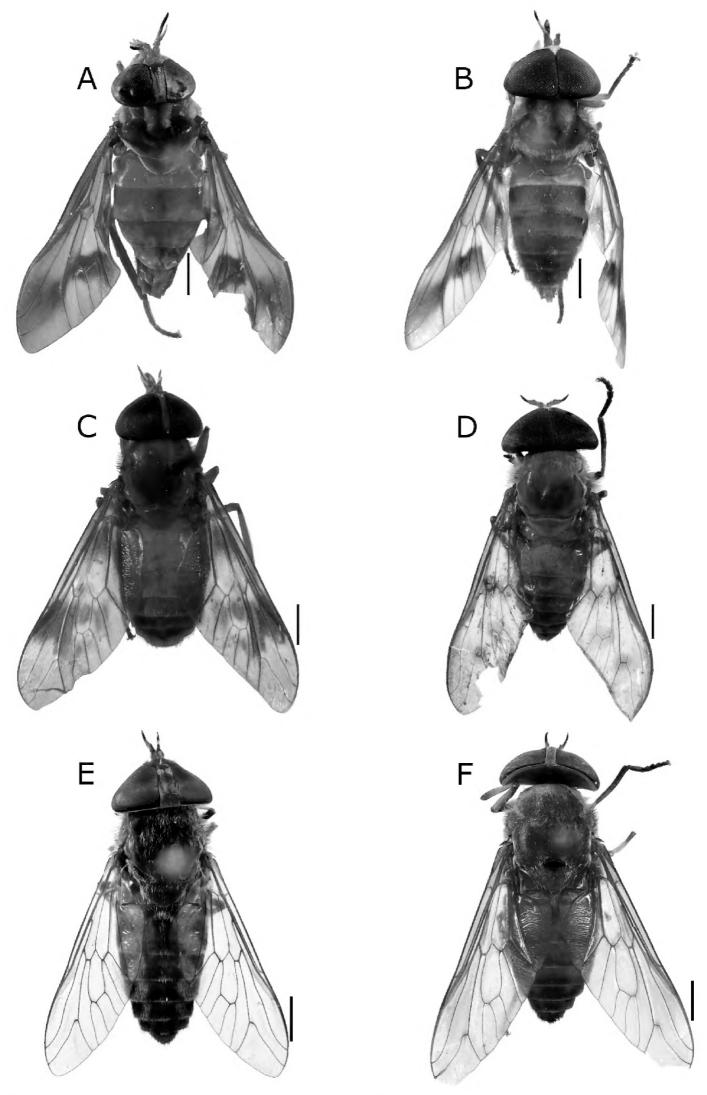


Figure 4. New records of Tabanidae from Honduras **A, B** *Dichelacera submarginata* Lutz $(\capprox, \capprox, \ca$

Material examined. Honduras: 1 ♂, Santa Bárbara, El Volcán, Trinidad, 15°08'02"N, 88°18'01"W, 1320 m, 26.vi.2000. R. Cordero and J. Torres leg.; EAPZ35.149.

TABANINI

Tabanus claripennis (Bigot, 1892)

Figure 4E

Distribution. Previously known from the West Indies, Costa Rica to Paraguay, Brazil, Argentina, and Chile (Coscarón and Papavero 2009).

Material examined. Honduras: 7 ♀♀, Francisco Morazán, El Zamorano, EAP, 14°01'N, 87°00'W, 5–29.vii.2020, H-trap, R. Argueta leg.; EAPZ43.572. 1♂, Choluteca, 6.7 km SE Santa Ana de Yusguare, 13°15'37"N, 87°04'40"W, 8.ix.1999, R. Cave and J. Torres leg.; EAPZ43.570.

Tabanus nebulosus De Geer, 1776

Figure 4F

Distribution. Previously known from Belize (Coscarón and Papavero 2009), Costa Rica (Fairchild 1961), Colombia, Venezuela, Trinidad, Surinam, Brazil, Bolivia, Paraguay, Barbados, and Argentina (Coscarón and Papavero 2009; Henriques 2016).

Material examined. Honduras: 2 ♀♀, Francisco Morazán, El Zamorano EAP, 14°01'N, 87°00'W, 850 m, v–vii, Estudiante EAPZ leg.; EAPZ75.022. 1♀, Francisco Morazán, El Zamorano EAP, 14°01'N, 87°00'W, 850 m, 31.v.2019, L. Moreno leg.; EAPZ75.023.

Key to the subfamilies, tribes, and genera of Tabanidae from Honduras

Modified from Fairchild (1969) and Burger (2009).

1	Hind tibiae without paired terminal spurs or spines; TABANINAE	5
_	Hind tibiae with paired terminal spurs or spines, spines rarely absent or dif	-
	ficult to see	2
2	Third antennal segment with 7 or 8 distinct flagellomeres; tergite 9 undi	_
	vided; PANGONIINAE	3
_	Third antennal segment with no more than 5 distinct flagellomeres; tergite 9)
	divided; CHRYSOPSINAE	5
3	Eyes bare; frons with ridge-like callus, which may be bare or tomentose	•
	PANGONIINI Esenbeckia Rondan	i
_	Eyes pilose; frons flat, without any sort of callus; SCIONINI	í
4	Cell m ₃ closed at wing margin	r
_	Cell m ₃ open at wing margin	r

5	Wings with dark crossband (Fig. 2A), crossband absent at times; eyes in life
	with pattern of dots and bars
_	Wings hyaline or cloudy on cross veins or elsewhere, without distinct cross-
6	band; eye pattern in life irregularly speckled
O	on adjoining costa; if setae on basicosta as dense as on costa, then vestiges of
	ocelli present; DIACHLORINI7
_	Basicosta with numerous strong setae, setae equal in size and density to
	those on adjoining costa, if setae sparse, then without vestiges of ocelli;
	TABANINI
7	Third antennal segment with strong dorso-basal tooth or forward-pointing
•	spine that often reaches to or beyond end of first flagellomere
_	Third antennal segment usually at most with acute dorso-basal angle12
8	Eyes densely pilose; antennal tooth reaching beyond apex of first flagellomere;
	proboscis longer than maxillary palpi; maxillary palpi slender, generally
	exceeding antennae; labella short, membranous; callus club shaped, much
	narrower than frons; wings with diffuse dark discal marking
_	Eyes bare; other characters variable9
9	Stout species; body sometimes hairy and beelike; foretibiae usually inflated;
	long hair fringes on at least hind tibiae; maxillary palpi inflated; antennae
	short, stout, with dorsal tooth extending beyond apex of first flagellomere;
	labella shiny and sclerotized10
_	Slender species; all tibiae slender; rest of characters not as above11
10	Abdomen green or greenish, sparsely covered with hairs; hind tibial fringe
	moderate in length; all tibiae slender; wings hyaline, sometimes yellowish;
	not resembling bees
_	Abdomen not greenish, densely hirsute; hind tibial fringe long; at least
	foretibia inflated; wings variable, never entirely hyaline or uniformly tinted,
	generally with black or contrasting pattern; body often resembling bees (see
11	Turcatel et al. 2010)
11	Basal callus thin, ridge-like, narrower than frons; eyes unicolored, bright green in life, rarely bicolored or with faint median line; mesoscutum uni-
	colored or weakly striped, not transversely banded
_	Basal callus as wide as frons; eyes banded or unicolorous blackish in life;
	mesoscutum often transversely banded
12	Subcallus, and usually first antennal segment, greatly inflated and shiny; third
	antennal segment long and slender, with obtuse dorso-basal angle; tibiae slen-
	der or slightly incrassate; wings black or partly so, with apex sharply hyaline,
	apical half of vein R ₄ bent sharply forward; maxillary palpi moderately slen-
	der, tomentose; clypeus tomentose
_	Without above combination of characters13

13	Tibiae, especially first two pairs, greatly inflated; subcallus, clypeus, and gena bare; maxillary palpi shiny and flattened; wings black at base, at least to ends
	of cells br and bm; labella membranous
_	Tibiae not or but slightly inflated; without above combination of characters15
14	Large, shiny bluish-black species; wings black from base to middle of cell d
_	Small species, mesoscutum, and often abdomen, with metallic brassy or
	greenish scale-like hairs; wings black from base to beyond end of cell d, with
	hyaline triangle in cells m ₃ and cua ₁
15	Mesopleura shiny or pearly tomentose in contrast to rest of pleura; wings usu-
	ally with dark subapical marking
_	Mesopleura not shiny or pearly tomentose, not contrasting with other pleural
	sclerites; wings without dark subapical marking16
16	Basal callus absent
_	Basal callus present, reduced at times
17	Labella sclerotized; frons narrow, generally over 5 times as long as its basal
	width; eyes in life unicolored, unbanded; dorsal angle on third antennal seg-
	ment strong
_	Labella membranous; frons generally less than 4 times as long as its basal width;
	eyes in life usually banded; dorsal angle of third antennal segment variable 18
18	Eyes bare, with at least 2 transverse bands in life; mostly small species with
	moderately broad frons often with median dark-haired patch; callus rounded
	or square, generally as wide as frons
_	Eyes pilose or bare, with at most 1 dark median, generally unicolored, rarely
	bicolored; rest of characters not as above
19	Vertex with well-marked tubercle and/or with clear vestiges of ocelli; eyes
	bare; frons narrow; basal callus club-shaped or ridge-like
_	Vertex without tubercle or clear vestiges of ocelli, slightly raised shiny or dis-
	colored tubercle rarely present; if tubercle present, then eyes pilose, or frons
• •	broad, or basal callus rounded
20	Wings with extensive dark pattern not consisting of spots on cross veins; if
	wings apparently unmarked, then thorax prominently striped, or frons ex-
	ceedingly narrow and callus thread-like
_	Wings hyaline, tinted, or with dark pattern consisting primarily of dark spots
21	around cross veins
21	Wings hyaline or evenly tinted, with costal cell often darker, but never with api-
	cal clouds or spots on cross veins; frontal callus clavate or ridge-like; abdomen
	black or brown, nearly always with transverse bands at least on fourth segment,
	rarely otherwise; appendix on fork of vein R ₄ absent <i>Leucotabanus</i> Lutz
_	Wing with clouds on at least discal cross veins, often with apical infuscation,
	if entirely hyaline or tinted, then abdomen and thorax not as above; frontal
	callus variable; wings often with appendix on fork of vein R ₄
	Siypommisa Enderiein

22	Vertex with small, rounded, sometimes indistinct, tubercle; eyes of female
	usually pilose, densely so on males; wings with all cross veins prominently
	spotted
_	Vertex rarely with tubercle; without above combination of characters

Acknowledgements

We are grateful to the reviewers, Daniel Carmo and Mauren Turcatel, as well as the editor, Torsten Dikow, for critically reading the manuscript and contributing to improve the quality of this paper. We thank Milena Agila for taking the photographs and arranging the plates for publication. We also thank Ronel Argueta for his help constructing and maintaining the traps at Zamorano.

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